

Loss of Muscle Mass During Chemotherapy Is Predictive for Poor Survival of Patients With Metastatic Colorectal Cancer

Susanne Blauwhoff-Buskermolen, Kathelijnn S. Versteeg, Marian A.E. de van der Schueren, Nicole R. den Braver, Johannes Berkhof, Jacqueline A.E. Langius, and Henk M.W. Verheul

Susanne Blauwhoff-Buskermolen, Kathelijnn S. Versteeg, Marian A.E. de van der Schueren, Nicole R. den Braver, Johannes Berkhof, Jacqueline A.E. Langius, and Henk M.W. Verheul, VU University Medical Center, Amsterdam; Marian A.E. de van der Schueren, HAN University of Applied Sciences, Nijmegen; and Jacqueline A.E. Langius, The Hague University of Applied Sciences, The Hague, the Netherlands.

Published online ahead of print at www.jco.org on February 22, 2016.

Authors' disclosures of potential conflicts of interest are found in the article online at www.jco.org. Author contributions are found at the end of this article.

Corresponding author: Henk M.W. Verheul, MD, PhD, Department of Medical Oncology, VU University Medical Center, P.O. Box 7057, 1007 MB Amsterdam, the Netherlands; e-mail: h.verheul@vumc.nl.

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0732-183X/15/3499-1/\$20.00

DOI: 10.1200/JCO.2015.63.6043

A B S T R A C T

Purpose

Low muscle mass is present in approximately 40% of patients with metastatic colorectal cancer (mCRC) and may be associated with poor outcome. We studied change in skeletal muscle during palliative chemotherapy in patients with mCRC and its association with treatment modifications and overall survival.

Patients and Methods

In 67 patients with mCRC (mean age \pm standard deviation, 66.4 ± 10.6 years; 63% male), muscle area (square centimeters) was assessed using computed tomography scans of the third lumbar vertebra before and during palliative chemotherapy. Treatment modifications resulting from toxicity were evaluated, including delay, dose reduction, or termination of chemotherapy. Multiple regression analyses were performed for the association between change in muscle area and treatment modification and secondly overall survival.

Results

Muscle area of patients with mCRC decreased significantly during 3 months of chemotherapy by 6.1% (95% CI, -8.4 to -3.8 ; $P < .001$). Change in muscle area was not associated with treatment modifications. However, patients with muscle loss during treatment of 9% or more (lowest tertile) had significantly lower survival rates than patients with muscle loss of less than 9% (at 6 months, 33% v 69% of patients alive; at 1 year, 17% v 49% of patients alive; log-rank $P = .001$). Muscle loss of 9% or more remained independently associated with survival when adjusted for sex, age, baseline lactate dehydrogenase concentration, comorbidity, mono-organ or multiorgan metastases, treatment line, and tumor progression at first evaluation by computed tomography scan (hazard ratio, 4.47; 95% CI, 2.21 to 9.05; $P < .001$).

Conclusion

Muscle area decreased significantly during chemotherapy and was independently associated with survival in patients with mCRC. Further clinical evaluation is required to determine whether nutritional interventions and exercise training may preserve muscle area and thereby improve outcome.

J Clin Oncol 34. © 2016 by American Society of Clinical Oncology

INTRODUCTION

Colorectal cancer (CRC) is the third most common cancer in the world, with nearly 1.4 million new cases diagnosed in 2012.¹ Approximately one-fifth of patients are diagnosed with synchronous metastatic disease (mCRC), and almost 50% of patients will eventually develop metastatic disease after their primary diagnosis. A large number of patients with mCRC are overweight or obese, with prevalence ranging between 45% and 53%.²⁻⁴ Low muscle mass is present in approximately 40% of patients, and not only in those

who are underweight.³ Low muscle mass in combination with a body mass index (BMI) greater than 30 kg/m² is referred to as sarcopenic obesity and was a prognostic indicator of worse overall survival in patients with pancreatic cancer⁵ and patients with solid tumors of the respiratory or GI tract.⁶

Causes of low muscle mass may be found in a combination of reduced food intake, low physical activity, and abnormal metabolism.⁷ Abnormal metabolism was found to be the most important contributor to low muscle mass in progressive end-stage disease⁸⁻¹⁰ and is referred to as cachexia. Reversing muscle loss in patients with

cancer is difficult, because cachexia is caused by tumor activity and released cytokines. Nevertheless, during active anticancer treatment, there might be a window of opportunity for nutritional support and exercise programs.

Cross-sectional studies have shown that low muscle mass is associated with poor survival and increased treatment toxicity in patients with CRC undergoing treatment with chemotherapy.^{2,3,11-13} However, longitudinal studies on change in muscle mass during chemotherapy for mCRC are lacking. The aim of this prospective study was to evaluate muscle mass change using single computed tomography (CT) images of the abdominal region (L3) in relation to chemotherapy toxicity and survival in patients with mCRC. Second, baseline muscle measurements in relation to clinical benefit and toxicity resulting from chemotherapy were evaluated.

PATIENTS AND METHODS

Data were prospectively collected in a study on (pre)cachexia, in which nutritional and biochemical features from patients with advanced cancer were evaluated before start of chemotherapy at the Vrije Universiteit Medical Center (Amsterdam, the Netherlands). A first interim analysis of these data was published in 2014.¹⁴ Patients were invited to enter the study before start of chemotherapy. Inclusion criteria were: adult patients with stage III to IV lung cancer, stage IV breast cancer, stage IV prostate cancer, or stage IV CRC for whom chemotherapy was planned. Exclusion criteria were: systemic treatment in the past month and clinically overt ascites or serious pitting edema.

Measurements of nutritional and biochemical features were performed at one time point: before start of chemotherapy treatment. For the current subgroup analyses, all consecutive patients with mCRC, recruited between October 2011 and January 2014, were selected. CT scans made for diagnostic purposes, at baseline, and during treatment with chemotherapy (according to standard evaluation schemes) were used to evaluate change in muscle mass during treatment.

The research protocol was approved by the Medical Ethics Committee of the Vrije Universiteit Medical Center Amsterdam, and the study was performed in accordance with the ethical standards described in the 1964 Declaration of Helsinki. Written informed consent was obtained from all participants.

Measurements

Skeletal muscle measurement. Skeletal muscle area (square centimeters) was measured with SliceOmatic software (version 5.0; Tomovision, Magog, Quebec, Canada) using routine CT scans conducted for diagnostic purposes. We used two CT scans: one at baseline (before start of chemotherapy) and one during treatment, according to standard evaluation schemes. Median time between the two scans was 78 days (interquartile range, 67 to 92 days). The third lumbar vertebra (L3) was used as a standard landmark, because this correlates best with whole-body muscle mass^{15,16}; the first image extending from L3 to the iliac crest was chosen to measure total muscle cross-sectional area. The L3 region contains psoas, paraspinal muscles, and the abdominal wall muscles. These muscles were identified on the basis of their anatomic features by two trained researchers (S.B.-B. and N.R.d.B.). The structures of those specific muscles were quantified on the basis of pre-established thresholds of Hounsfield units (−29 to 150) of skeletal muscle tissue.¹⁷ Cross-sectional areas (square centimeters) of the sum of all these muscles were computed by adding tissue pixels and multiplying by the pixel surface area for each patient at each time point. We found a mean coefficient of variation between observers of 0.6% for skeletal muscle area in a random sample of 20 patients, which is regarded to be low.¹⁸ To estimate total-body skeletal muscle mass, the regression equation of Shen et al¹⁵ was used. Changes

between the first and second scans were calculated as a rate of change per 3 months, meaning change per first evaluation, to compare with available literature.^{19,20} Relative muscle change per 3 months was categorized into tertiles of muscle change: tertile one, highest muscle loss up to 9%; tertile two, muscle loss of 9% to 1.5%; and tertile three, muscle loss of 1.5% until highest gain in muscle.

Muscle density (MD) was measured using the muscle radiation attenuation rate (in Hounsfield units) because of its prognostic value in patients with cancer.^{21,22} Skeletal muscle index (SMI) was calculated as the ratio of skeletal muscle area (square centimeters) divided by height (meters).² Body height was measured using a stadiometer; the patient stood barefoot, and height was determined to the nearest centimeter. Sex- and BMI-specific cutoff values of low SMI and low MD according to Martin et al²³ were used to define patients with normal and low values.

Body weight. Baseline body weight was measured within 0.2 kg on a calibrated scale (Seca type 888), and self-reported weight loss in the past 6 months was noted. Body weight during chemotherapy was obtained from medical records within 2 weeks of the date of the second CT scan. A correction factor for clothes or clothes and shoes was made by deducting weight by 1.6 and 2.0 kg for men and 1.0 and 1.3 kg for women, respectively.²⁴ To compare with change in muscle mass, we calculated the rate of change (percent) standardized to 3 months. BMI was calculated as the ratio of body weight (kilograms) divided by height (meters).²

Treatment Modifications

Treatment modifications that were taken into account consisted of delay, dose reduction, or discontinuation of chemotherapy because of reported toxicity in the period between the two CT scans. Other modifications, such as delay because of patient preference or vacation, were not taken into consideration. Also, the number of treatment modifications was not taken into account, because treatment modifications were measured on a dichotomous scale (present or absent). Information on treatment modifications was obtained from medical records.

Survival

Total follow-up time was 3.5 years. Survival time was defined as time from inclusion in the study until death or last consultation in the hospital.

Covariates

The following variables were obtained from medical records and used in statistical analyses as covariates, because they carry prognostic value: sex, age, treatment line (\geq second v first, counted as consecutive chemotherapy line on the basis of the fact that a new type of chemotherapy was introduced), start dose of chemotherapy (normal or decreased), WHO performance status (≥ 2 v 0 or 1), Charlson comorbidity index²⁵ (≥ 1 v 0), serum lactate dehydrogenase (LDH; measured within 4 weeks before start of chemotherapy), severity of disease (multiorgan v monoorgan metastases), and outcome of first evaluation CT scan (progression v stable disease or regression of the tumor).

Statistics

Statistical analyses were performed using SPSS software for Windows (version 20.0; SPSS, Chicago, IL). Descriptive statistics (count [percent], mean \pm standard deviation, or median and interquartile range, as appropriate) were used to describe the study sample.

Paired-sample *t* tests were conducted to assess changes in skeletal muscle area, MD, and body weight. Spearman's correlation coefficient was used to assess correlation between body weight change and muscle area change.

Logistic regression analyses were used to test associations between treatment modification and muscle measurement (baseline SMI, low v normal [reference]; baseline MD, low v normal [reference]; and change in muscle area per percent). In multiple regression analyses, adjustments were made for age, sex, comorbidity score, start dose, treatment line, and WHO performance status.

Kaplan-Meier curves for overall survival were constructed separately for patients in the first, second, and third tertiles of relative muscle change. Differences between the curves were evaluated by log-rank tests. Six-month and 1-year survival proportions were also calculated. Cox proportional hazards analyses were performed to test associations between overall survival and muscle measurements (baseline SMI, low ν normal [reference]; baseline MD, low ν normal [reference]; and change in muscle area per percent). In multiple regression analyses, adjustments were made for age, sex, baseline serum LDH level, comorbidity score, severity of disease, and treatment line. For analyses including change in muscle mass during chemotherapy, an extra adjustment was performed for tumor progression at first evaluation by CT scan (progression ν response or stable disease [reference]) as a potential confounder for rate of muscle loss and survival. Furthermore, overall survival times were calculated with the second CT scan as the starting point for these analyses. $P \leq .05$ was considered significant for all analyses.

RESULTS

Sixty-seven patients with mCRC and evaluable CT scans at baseline were included. For longitudinal analyses, four patients were excluded because they had died ($n = 2$) or because they had no CT scans for other reasons ($n = 2$) at the time of evaluation.

Baseline Patient and Treatment Characteristics

The mean age of patients (\pm standard deviation) was 66.4 ± 10.6 years, and 63% of patients were male. Eighty-two percent of patients had multiorgan metastases, and 78% received first-line

Characteristic	No. (%)
Age, years	
Mean	66.4
SD	10.6
Sex	
Male	42 (63)
Female	25 (37)
Metastases	
Liver only	12 (18)
Multiorgan	55 (82)
Treatment line	
First	52 (78)
\geq Second	15 (23)
Starting dose of chemotherapy	
Full	52 (78)
Reduced	15 (22)
Chemotherapy type	
CAPOX (\pm bevacizumab)	44 (66)
FU plus oxaliplatin (\pm bevacizumab)	5 (7)
Capecitabine plus irinotecan	2 (3)
Capecitabine plus bevacizumab	1 (1)
Capecitabine monotherapy	5 (7)
Irinotecan monotherapy	10 (15)
WHO performance status	
0	27 (40)
1	35 (52)
≥ 2	5 (8)
Charlson comorbidity index	
0	45 (67)
1-2	19 (28)
≥ 3	3 (5)

Abbreviations: CAPOX, capecitabine plus oxaliplatin; FU, fluorouracil; SD, standard deviation.

Table 2. Nutritional Status at Baseline (N = 67)

Variable	No. (%)
Weight change in 6 months before baseline	
Gain > 5%	6 (9)
Stable	39 (58)
Loss 5% to 10%	15 (22)
Loss > 10%	7 (10)
BMI, kg/m ²	
< 20	4 (6)
20-24.9	21 (31)
25-29.9	37 (55)
> 30	5 (8)
SMI < reference value*	38 (57)
MD < reference value*	43 (64)
Sarcopenic obesity†	1 (2)

Abbreviations: BMI, body mass index; MD, muscle density; SMI, skeletal muscle index.
 *Sex- and BMI-specific cutoff values according to Martin et al.²³
 †BMI ≥ 30 kg/m² and low SMI according to Martin et al.²³

chemotherapy. Patients received standard systemic treatment for CRC; the majority ($n = 44$; 66%) received a combination of capecitabine plus oxaliplatin with or without bevacizumab (CAPOX B or CAPOX, respectively; Table 1).

Critical weight loss of greater than 5% in 6 months before start of chemotherapy was prevalent in 33% of patients. At baseline, more than half of the patients were overweight (55%) or obese (8%), and 57% had low SMI. Sarcopenic obesity was present in one patient (Table 2).

Changes in Muscle Mass and Body Weight

Skeletal muscle area decreased, in both men and women, by 6.1% (95% CI, -8.4 to -3.8 ; $P < .001$) in 3 months, corresponding to 1.7 kg in men and 1.1 kg in women. The proportion of patients with low SMI increased from 57% at baseline to 70% at the second CT scan (Table 3).

Body weight decreased significantly in women by 4.4% (95% CI, -8.5 to -0.4 ; $P = .031$), corresponding with -2.9 kg ($n = 18$), but not in men (change, -3.3% ; 95% CI, -8.1 to 1.5 ; $P = .167$). Change in muscle area correlated significantly with change in body weight for men, but not for women (Spearman's rho for men, 0.34; $P = .050$; for women, 0.36; $P = .140$). MD decreased in men by 5.8% (95% CI, -11.5 to -0.03 ; $P = .049$) but remained stable in women (-0.3% ; 95% CI, -11.7 to 11.1 ; $P = .957$; Table 3).

Treatment Modifications

Treatment modifications resulting from toxicity occurred in 29 patients (43%), including delay of treatment in 13 patients (19%), dose reduction in 18 patients (27%), and discontinuation of treatment in six patients (9%). SMI and MD at baseline were not associated with treatment modifications (SMI: odds ratio [OR], 1.01; 95% CI, 0.35 to 2.91; $P = .99$; MD: OR, 1.43; 95% CI, 0.44 to 4.63; $P = .555$). In the more homogenous group of patients undergoing first-line treatment with CAPOX, no association was found either ($n = 35$; SMI: OR, 0.64; 95% CI, 0.07 to 5.71; $P = 0.692$; MD: OR, 1.21; 95% CI, 0.15 to 9.96; $P = .857$). A change in skeletal muscle area during treatment was not associated with treatment modifications in the total cohort (OR per 1% decrease,

Table 3. Muscle Area and Density During Chemotherapy (n = 63)

Variable	First CT Scan		Second CT Scan		Change			Change Per 3 Months			Relative Change Per 3 Months (%)		
	Mean	SD	Mean	SD	Mean	95% CI	P	Mean	95% CI	P	Mean	95% CI	P
Muscle area, cm ²	138.6	32.1	131.9	31.7	-6.6	-8.9 to -4.3	< .001	-8.6	-11.8 to -5.3	< .001	-6.1	-8.4 to -3.8	< .001
Men	156.8	24.4	149.2	25.3	-7.6	-11.0 to -4.3	< .001	-9.7	-14.6 to -4.9	< .001	-6.2	-9.3 to -3.1	< .001
Women	108.9	17.5	103.9	17.9	-5.0	-8.0 to -1.9	.003	-6.6	-10.5 to -2.7	.002	-6.0	-9.5 to -2.5	.002
Muscle density, HU	33.8	8.1	32.8	8.8	-1.1	-2.4 to 0.2	.106	-1.6	-3.2 to 0.0	.045	-3.7	-9.2 to 1.8	.182
Men	35.2	7.5	33.6	9.0	-1.6	-3.2 to -0.1	.037	-2.0	-3.9 to -0.2	.031	-5.8	-11.5 to -0.03	.049
Women	31.6	8.8	31.5	8.4	-0.1	-2.6 to 2.3	.905	-0.9*	-4.0 to 2.1	.530	-0.3	-11.7 to 11.1	.957

Abbreviations: CT, computed tomography; HU, Hounsfield unit; SD, standard deviation.
*Skewed data: median, -0.3; interquartile range, -3.8 to 4.1.

1.03; 95% CI, 0.95 to 1.12; $P = 0.441$) or a homogenous group of patients undergoing first-line treatment with CAPOX (n = 34; OR, 1.07; 95% CI, 0.95 to 1.22; $P = .274$).

Survival

Median overall survival was 17.5 months (95% CI, 13.3 to 21.7) for patients receiving first-line chemotherapy and 8.5 months (95% CI, 4.4 to 12.6) for patients receiving second-line chemotherapy or beyond. Survival curves were significantly different for the tertiles of muscle change (log-rank $P = .005$; Fig 1). In pairwise comparison, the survival curve of tertile one ($\geq 9\%$ muscle loss) was significantly different from the survival curve of tertile two (muscle loss of 1.5% to 9%; log-rank $P = .007$) and also from the survival curve of tertile three (muscle loss of 1.5% to highest gain in muscle; log-rank $P = .009$). However, the survival curve of tertile two was not significantly different from the survival curve of tertile three (log-rank $P = .961$); therefore tertiles two and three were pooled (cutoff, 9% muscle loss).

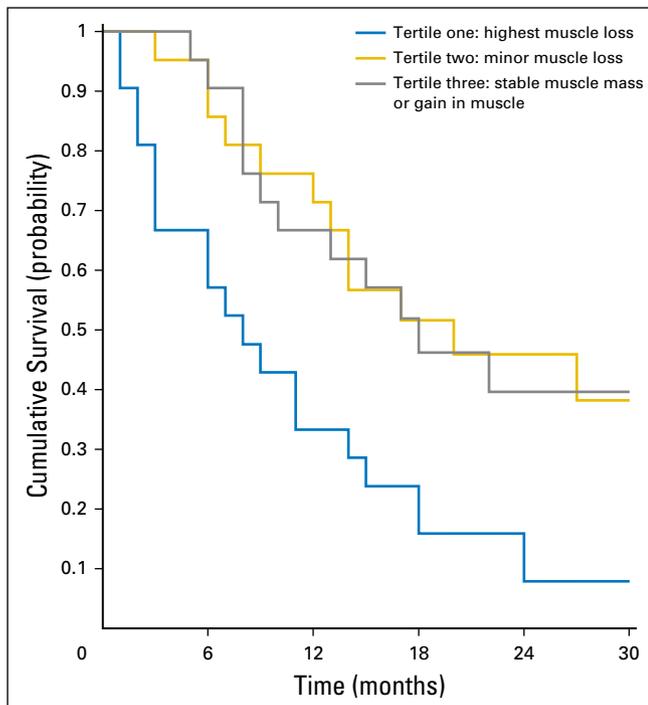


Fig 1. Kaplan-Meier curves for tertiles of muscle change.

Patients with muscle loss of 9% or more (tertile one) during chemotherapy had significantly lower overall survival rates than patients with muscle loss of less than 9% (tertiles two and three); at 6 months, 33% versus 69% of patients were alive, and at 1 year, 17% versus 49% of patients were alive (log-rank $P = .001$). Muscle loss of 9% or more remained independently associated with shorter survival when adjusted for sex, age, baseline LDH concentration, comorbidity, mono-organ or multiorgan metastases, treatment line, and tumor progression at first evaluation by CT scan (hazard ratio, 4.47; 95% CI, 2.21 to 9.05; $P < .01$). Low MD at baseline was associated with shorter survival after adjustment for covariates (hazard ratio, 2.38; 95% CI, 1.16 to 4.87; $P = .018$). Low SMI at baseline was not associated with survival (Table 4).

DISCUSSION

This is the first study to our knowledge to present a change in muscle area during palliative chemotherapy for patients with mCRC. Patients with a decrease in muscle area during chemotherapy of 9% or more—the lowest tertile of patients—had significantly lower overall survival rates than patients with a decrease of less than 9%, independent of known prognostic covariates.

Previously, a rapid decrease in skeletal muscle mass was shown in patients with CRC in the last year of their life.^{8,9} In our study, patients lost skeletal muscle during active anticancer treatment. The rate of muscle loss was comparable to that of patients with cholangiocarcinoma or esophageal cancer during treatment.^{19,20} Compared with muscle loss that occurs in normal aging, patients with cancer experience a 24-fold more rapid loss of muscle mass: 6% muscle loss in 3 months (Table 2) versus 1% per year.²⁶ Overall consequences of muscle atrophy in humans have been found to be devastating: increased risk of falling,²⁷ diminished self-reliance,²⁸ decreased quality of life, increased risk of treatment-related complications, and shorter survival.²⁹

Although previous studies have described an association between reduced muscle mass before treatment and treatment modifications,^{2,11} we did not observe this association. A possible explanation for this may be found in the heterogeneity regarding treatment regimens and follow-up time; six different treatment regimens were recorded, and follow-up time depended on the timing of the second CT scan. Prado et al¹¹ and Barrett et al² looked at homogeneous groups of patients with mCRC receiving one type of chemotherapy and specific follow-up time (eg, number of

Table 4. Associations Between SMI, MD, and Muscle Change With Overall Survival

Variable	Unadjusted		Adjusted	
	HR (95% CI)	P	HR (95% CI)	P
Low SMI at baseline (N = 67)	1.30 (0.72 to 2.35)	.390	1.65 (0.85 to 3.18)*	.138
Low MD at baseline (N = 67)	1.36 (0.74 to 2.50)	.321	2.38 (1.16 to 4.87)*	.018
≥ 9% muscle loss (n = 63)	2.69 (1.45 to 5.01)	.002	4.47 (2.21 to 9.05)†	< .001

Abbreviations: HR, hazard ratio; MD, muscle density; SMI, skeletal muscle index.

*Adjusted for sex, age, lactate dehydrogenase concentration, comorbidity, metastases, and chemotherapy line.

†Adjusted for sex, age, lactate dehydrogenase concentration, comorbidity, metastases, chemotherapy line, and tumor progression at first evaluation by computed tomography scan.

cycles). Nevertheless, in post hoc analyses, no association between skeletal muscle and treatment toxicity in a homogeneous group of patients treated with CAPOX in first-line chemotherapy was found. We hypothesize that muscle mass may not be related to bone marrow toxicity, one of the most important types of toxicity resulting from this treatment regimen.³⁰ Furthermore, peripheral neuropathy is a type of toxicity that might be related to muscle loss but has been regarded as a more long-term type of toxicity and therefore may not yet be present in the first 3 months of treatment.

Although there was an association with low MD at baseline (in multiple regression analysis only), low SMI at baseline was not associated with survival in the evaluated group of patients with mCRC. This is in contrast with some^{5,6,12,31,32} but not all previous findings. Stene et al³³ described comparable results in non-small-cell lung cancer, and Antoun et al²² also reported comparable results in patients with melanoma. One possible explanation might be found in the phenomenon of sarcopenic obesity; patients with sarcopenic obesity had worse prognosis compared with patients without sarcopenic obesity in earlier studies.^{5,6} Although the number of obese patients in our study was too small for subgroup analyses, we found that the prevalence of low muscle mass in obese patients in our study was comparable to the prevalence found in the study by Prado et al⁶: 20% (one of five) versus 15% (38 of 250), respectively. Muscle loss during chemotherapy was associated with worse overall survival, independent of important prognostic covariates. This is a new finding and raises the question whether interventions aimed at preservation of muscle mass during treatment are effective in improving outcome. One example of a promising intervention might be increasing physical activity during treatment, because this has been proven to improve muscle strength in cancer survivors³⁴ and in patients during chemotherapy.³⁵ Another promising intervention might be nutritional counseling, with consumption of high amounts of protein during the day³⁶ with high-enough thresholds per meal,³⁷ because this has also been proven to increase protein synthesis in healthy elderly populations. Potentially, the two interventions may be optimal when combined.³⁸ The use of ghrelin agonists for

the treatment of cancer-associated anorexia could also be considered.³⁹ Furthermore, future studies should consider recruiting patients beyond those with severe weight loss or cachexia, because we have shown that muscle loss during treatment was a predictor of poor survival independent of the amount of muscle mass at baseline.

Alternatively, causes of muscle loss could be studied more in depth to find rational intervention possibilities. For example, the intervention strategy will be different when anorexia is the most important cause of muscle loss than when cachexia is the most important cause of muscle loss. To study this, a larger number of patients is required to also evaluate food intake and energy expenditure before and during chemotherapy.

In conclusion, significant muscle loss occurred in patients with mCRC during chemotherapy. Muscle loss of 9% or more during chemotherapy was independently associated with poorer survival. Future studies should investigate causes of muscle loss and whether interventions may attenuate or improve muscle mass during treatment and may lead to improvement in clinical outcomes such as survival.

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Disclosures provided by the authors are available with this article at www.jco.org.

AUTHOR CONTRIBUTIONS

Conception and design: Susanne Blauwhoff-Buskermol, Marian A.E. de van der Schueren, Jacqueline A.E. Langius, Henk M.W. Verheul
Collection and assembly of data: Susanne Blauwhoff-Buskermol, Kathelijn S. Versteeg, Nicole R. den Braver
Data analysis and interpretation: All authors
Manuscript writing: All authors
Final approval of manuscript: All authors

REFERENCES

1. World Health Organization International Agency for Research on Cancer: GLOBOCAN 2012: Estimated Cancer Incidence, Mortality and Prevalence Worldwide in 2012. <http://globocan.iarc.fr>

2. Barret M, Antoun S, Dalban C, et al: Sarcopenia is linked to treatment toxicity in patients with metastatic colorectal cancer. *Nutr Cancer* 66: 583-589, 2014

3. Thoresen L, Frykholm G, Lydersen S, et al: Nutritional status, cachexia and survival in patients with advanced colorectal carcinoma: Different

assessment criteria for nutritional status provide unequal results. *Clin Nutr* 32:65-72, 2013

4. Barret M, Malka D, Aparicio T, et al: Nutritional status affects treatment tolerability and survival in metastatic colorectal cancer patients: Results of an AGEO prospective multicenter study. *Oncology* 81: 395-402, 2011

5. Tan BHL, Birdsall LA, Martin L, et al: Sarcopenia in an overweight or obese patient is an adverse prognostic factor in pancreatic cancer. *Clin Cancer Res* 15:6973-6979, 2009
6. Prado CMM, Lieffers JR, McCargar LJ, et al: Prevalence and clinical implications of sarcopenic obesity in patients with solid tumours of the respiratory and gastrointestinal tracts: A population-based study. *Lancet Oncol* 9:629-635, 2008
7. Fearon K, Strasser F, Anker SD, et al: Definition and classification of cancer cachexia: An international consensus. *Lancet Oncol* 12:489-495, 2011
8. Lieffers JR, Mourtzakis M, Hall KD, et al: A viscerally driven cachexia syndrome in patients with advanced colorectal cancer: Contributions of organ and tumor mass to whole-body energy demands. *Am J Clin Nutr* 89:1173-1179, 2009
9. Prado CM, Sawyer MB, Ghosh S, et al: Central tenet of cancer cachexia therapy: Do patients with advanced cancer have exploitable anabolic potential? *Am J Clin Nutr* 98:1012-1019, 2013
10. Wallengren O, Iresjö B, Lundholm K, et al: Loss of muscle mass in the end of life in patients with advanced cancer. *Support Care Cancer* 23:79-86, 2015
11. Prado CMM, Baracos VE, McCargar LJ, et al: Body composition as an independent determinant of 5-fluorouracil-based chemotherapy toxicity. *Clin Cancer Res* 13:3264-3268, 2007
12. van Vledder MG, Levolger S, Ayez N, et al: Body composition and outcome in patients undergoing resection of colorectal liver metastases. *Br J Surg* 99:550-557, 2012
13. Malietzis G, Aziz O, Bagnall NM, et al: The role of body composition evaluation by computerized tomography in determining colorectal cancer treatment outcomes: A systematic review. *Eur J Surg Oncol* 41:186-196, 2015
14. Blauwhoff-Buskermolen S, de van der Schueren MA, Verheul HM, et al: "Pre-cachexia": A non-existing phenomenon in cancer? *Ann Oncol* 25:1668-1669, 2014
15. Shen W, Punyanitya M, Wang Z, et al: Total body skeletal muscle and adipose tissue volumes: Estimation from a single abdominal cross-sectional image. *J Appl Physiol* (1985) 97:2333-2338, 2004
16. Mourtzakis M, Prado CMM, Lieffers JR, et al: A practical and precise approach to quantification of body composition in cancer patients using computed tomography images acquired during routine care. *Appl Physiol Nutr Metab* 33:997-1006, 2008
17. Mitsiopoulos N, Baumgartner RN, Heymsfield SB, et al: Cadaver validation of skeletal muscle measurement by magnetic resonance imaging and computerized tomography. *J Appl Physiol* (1985) 85:115-122, 1998
18. MacDonald AJ, Greig CA, Baracos V: The advantages and limitations of cross-sectional body composition analysis. *Curr Opin Support Palliat Care* 5:342-349, 2011
19. Awad S, Tan BH, Cui H, et al: Marked changes in body composition following neoadjuvant chemotherapy for oesophagogastric cancer. *Clin Nutr* 31:74-77, 2012
20. Prado CMM, Bekaii-Saab T, Doyle LA, et al: Skeletal muscle anabolism is a side effect of therapy with the MEK inhibitor: Selumetinib in patients with cholangiocarcinoma. *Br J Cancer* 106:1583-1586, 2012
21. Sabel MS, Lee J, Cai S, et al: Sarcopenia as a prognostic factor among patients with stage III melanoma. *Ann Surg Oncol* 18:3579-3585, 2011
22. Antoun S, Lanoy E, Iacovelli R, et al: Skeletal muscle density predicts prognosis in patients with metastatic renal cell carcinoma treated with targeted therapies. *Cancer* 119:3377-3384, 2013
23. Martin L, Birdsall LA, MacDonald N, et al: Cancer cachexia in the age of obesity: Skeletal muscle depletion is a powerful prognostic factor, independent of body mass index. *J Clin Oncol* 31:1539-1547, 2013
24. Frank E, Dunlop AL: What does a patient's outfit weight? *Fam Med* 32:595-596, 2000
25. Charlson ME, Pompei P, Ales KL, et al: A new method of classifying prognostic comorbidity in longitudinal studies: Development and validation. *J Chronic Dis* 40:373-383, 1987
26. Frontera WR, Hughes VA, Fielding RA, et al: Aging of skeletal muscle: a 12-yr longitudinal study. *J Appl Physiol* (1985) 88:1321-1326, 2000
27. Landi F, Liperoti R, Russo A, et al: Sarcopenia as a risk factor for falls in elderly individuals: Results from the iSIRENTE study. *Clin Nutr* 31:652-658, 2012
28. Janssen I, Heymsfield SB, Ross R: Low relative skeletal muscle mass (sarcopenia) in older persons is associated with functional impairment and physical disability. *J Am Geriatr Soc* 50:889-896, 2002
29. Rolland Y, Czerwinski S, Abellan Van Kan G, et al: Sarcopenia: Its assessment, etiology, pathogenesis, consequences and future perspectives. *J Nutr Health Aging* 12:433-450, 2008
30. Cassidy J, Clarke S, Diaz-Rubio E, et al: XELOX vs FOLFOX-4 as first-line therapy for metastatic colorectal cancer: NO16966 updated results. *Br J Cancer* 105:58-64, 2011
31. Prado CMM, Baracos VE, McCargar LJ, et al: Sarcopenia as a determinant of chemotherapy toxicity and time to tumor progression in metastatic breast cancer patients receiving capecitabine treatment. *Clin Cancer Res* 15:2920-2926, 2009
32. Sjöblom B, Grönberg BH, Benth JS, et al: Low muscle mass is associated with chemotherapy-induced haematological toxicity in advanced non-small cell lung cancer. *Lung Cancer* 90:85-91, 2015
33. Stene GB, Helbostad JL, Amundsen T, et al: Changes in skeletal muscle mass during palliative chemotherapy in patients with advanced lung cancer. *Acta Oncol* 54:340-348, 2015
34. Speck RM, Courneya KS, Mâsse LC, et al: An update of controlled physical activity trials in cancer survivors: A systematic review and meta-analysis. *J Cancer Surviv* 4:87-100, 2010
35. Courneya KS, Segal RJ, Mackey JR, et al: Effects of aerobic and resistance exercise in breast cancer patients receiving adjuvant chemotherapy: A multicenter randomized controlled trial. *J Clin Oncol* 25:4396-4404, 2007
36. Paddon-Jones D, Leidy H: Dietary protein and muscle in older persons. *Curr Opin Clin Nutr Metab Care* 17:5-11, 2014
37. Paddon-Jones D, Rasmussen BB: Dietary protein recommendations and the prevention of sarcopenia. *Curr Opin Clin Nutr Metab Care* 12:86-90, 2009
38. Fearon K, Arends J, Baracos V: Understanding the mechanisms and treatment options in cancer cachexia. *Nat Rev Clin Oncol* 10:90-99, 2013
39. Zhang H, Garcia JM: Anamorelin hydrochloride for the treatment of cancer-anorexia-cachexia in NSCLC. *Expert Opin Pharmacother* 16:1245-1253, 2015



AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Loss of Muscle Mass During Chemotherapy Is Predictive for Poor Survival of Patients With Metastatic Colorectal Cancer

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Susanne Blauwhoff-Buskermolen
No relationship to disclose

Kathelijn S. Versteeg
No relationship to disclose

Marian A.E. de van der Schueren
Speakers' Bureau: Nutricia

Nicole R. den Braver
No relationship to disclose

Johannes Berkhof
No relationship to disclose

Jacqueline A.E. Langius
Travel, Accommodations, Expenses: Nutricia

Henk M.W. Verheul
Honoraria: Boehringer Ingelheim
Consulting or Advisory Role: Boehringer Ingelheim (Inst)
Research Funding: Amgen (Inst), VHS (Inst), Immunovo BV (Inst), Roche (Inst)

Acknowledgment

We thank Vickie Baracos, PhD, and Nina Esfandiari from the Cross Cancer Institute, Edmonton, Alberta, Canada, for their help and advice regarding the analysis of muscle mass on computed tomography scans.